

## **Supplemental Information – Swift, Jody. L.**

### ***Candidates Academic and honorary degrees;***

- B.Sc. Chemistry (Honors)
- Ph.D. Chemistry, Biophysical Chemistry

### ***Candidates Current Occupation;***

- Director, Special Projects and Strategic Initiatives, Faculty of Applied Science Dean's Office

### ***Candidate's current and past academic, societal, governmental, or commercial offices held;***

- 2002 – 2004 VP Internal Affairs, Chemistry Student Society (CS<sub>2</sub>) – University of Saskatchewan
- 2013 – 2017 Co-Chair, Department of Medicine Research Advisory Committee – UBC
- 2015 – 2017 Advisory Committee Member – Clinical Research Professionals of BC (CRPBC)
- 2018 – current Employer Representative – APSC Joint Occupational Health & Safety Committee
- 2019 – current Member, Local Safety Team – School of Nursing

### ***Candidate's current professional or business interests;***

My current professional interests are centered around supporting effective organizational leadership, strategy, and planning. I work directly with the Dean and Associate Dean leadership team to identify, prioritize and implement new initiatives across the five schools and six departments which comprise the Faculty of Applied Science. The role oversees the team of Executive Assistants supporting the Dean and Associate Dean, as well as the Special Projects and Office Manager for the Faculty. Acting as a liaison between the Faculty and key stakeholder groups I am responsible to helping to communicate the priorities of the Faculty and develop key messaging consistent with the Faculty vision and mission. In this capacity I work closely with Directors of HR, Capital Projects, Marketing & Communications and Finance and Operations to develop and execute specific high value projects.

Key duties: planning and setting strategic priorities, development of strategic vision for the Faculty, liaise with key stakeholders and stakeholder groups, meeting facilitation, manage and oversee the conceptualization, planning and implementation of new projects and programs within the Faculty, assemble and lead project teams, identify key performance indicators, metrics & reporting, academic program development and implementation, accreditation and awards.

I have no outside business interests

### ***Candidate's list of publications;***

1. "Automatic Quality Assessment of Echocardiograms Using Convolutional Neural Networks: Feasibility on the Apical Four-Chamber View", *IEEE Transactions on Medical Imaging* **2017**. (36); 1221 - 1230
2. "Simultaneous Analysis of 2D Echo Views for Left Atrial Segmentation and Disease Detection", *IEEE Transactions on Medical Imaging* **2017**. (36); 40 - 50
3. "Automatic quality assessment of apical four-chamber echocardiograms using deep convolutional neural networks" *Progress in Biomedical Optics and Imaging - Proceedings of SPIE* **2017**. (10133)

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4. “Quantification of receptor tyrosine kinase activation and transactivation by G-protein- coupled receptors using spatial intensity distribution analysis (SpIDA)” *Methods in Enzymology* **2013**. (522); 109 – 31
5. “Spatial Intensity Distribution Analysis (SpIDA). A New Tool for Receptor Tyrosine Kinase Activation and Transactivation Quantification” *Methods in Cell Biology*. **2013**. (117); 1 - 19
6. “Ligand-induced Clustering of EGF Receptors: A Quantitative Study by Fluorescence Image Moment Analysis” *Biophys. Chem.* **2012**. (161); 50 – 53
7. “Quantification of Receptor Tyrosine Kinase Transactivation through Direct Dimerization and Surface Density Measurements in Single Cells” *Proc. Natl. Acad. Sci.* **2011**. 108 (17); 7016 – 7021
8. “Revealing Protein Oligomerization and Densities in Situ Using Spatial Intensity Distribution Analysis” *Proc. Natl. Acad. Sci.* **2011**. 108 (17); 7010 – 7015
9. “Fluorescence Microscopy Investigations of Ligand Propagation and Accessibility Under Adherent Cells.” *Biointerphases*. **2010**. 5 (4); 139 – 148
10. “Live-cell Microscopy – tips and tools” *Journal of Cell Science*. **2009**. (122); 753 – 767
11. “Effects of Various Small-Molecule Anesthetics on Vesicle Fusion: A Study Using Two-Photon Fluorescence Cross-Correlation Spectroscopy” *Journal of Physical Chemistry B*. **2009**. 113 (30); 10357 – 10366
12. “A Quantum Dot-labeled Ligand-receptor Binding Assay for G protein-coupled Receptors Contained in Minimally Purified Membrane Nanopatches” *Methods in Molecular Biology* (Clifton, N.J.). **2009**. 552:329 – 341
13. “Nanoparticles as Fluorescent Labels: Is Size All that Matters?” *Biophysical Journal*. **2008**. (95);865 – 876
14. “Two-Photon Excitation Fluorescence Cross-Correlation Assay for Ligand-Receptor Binding: Cell Membrane Nanopatches Containing the Human  $\mu$ -Opioid Receptor.” *Analytical Chemistry*. **2007**. (79); 6783 – 6791
15. “Fluorescence Correlation Spectroscopy Using Quantum Dots: Advances, Challenges and Opportunities.” *Physical Chemistry Chemical Physics*. **2007**. (9); 1870 – 1880
16. “A Two-photon Excitation Fluorescence Cross-correlation Assay for a Model Ligand Receptor Binding System Using Quantum Dots.” *Biophysical Journal*. **2005**. (90); 1396 – 1410
17. “Anesthetic Enhanced Membrane Fusion Examined Using Two-Photon Excitation Fluorescence Cross-correlation Spectroscopy”, *Journal of Physical Chemistry B* (**Cover Article**). **2004**. 108(30)